Evaluation of *Haemophilus parasuis* control in the nursery using vaccination and controlled exposure

Simone Oliveira, DVM, MS, PhD; Carlos Pijoan, DVM, MS, PhD; Robert Morrison, DVM, PhD, MBA

Summary

Objective: To evaluate the effectiveness of three control measures in reducing nursery mortality caused by *Haemophilus parasuis*, namely, a commercial vaccine, an autogenous vaccine, and controlled exposure using a low dose of live, virulent organisms.

Methods: The experiments were performed in a multi-farm system experiencing high nursery mortality (> 4.8%) due to *H parasuis* infection. In Experiment 1, pigs were vaccinated at weaning using a commercially available, one-dose *H parasuis* vaccine. In Experiment 2, pigs were vaccinated at processing and at weaning with an autogenous vaccine. In Experiment 3, pigs

were orally inoculated at processing using a bacterial suspension containing a total of 10^{5-6} colony forming units per mL of three *H parasuis* strains prevalent in the studied herd. Experiments 1 and 2 were repeated five times (5 consecutive weeks of production), and Experiment 3 was repeated nine times.

Results: Mortality in pigs exposed to H parasuis was lower (P < .001) than that in groups vaccinated with either commercial or autogenous vaccines.

Discussion: The lack of effect of parenteral vaccination may be due to lack of cross-protection between heterologous strains, timing of vaccination, and potential interference of maternally-derived immunity.

The efficacy of controlled exposure likely reflects the induction of homologous mucosal immunity preventing pathogen systemic invasion.

Implications: The use of controlled exposure of young pigs to the prevalent *H* parasuis strains involved in nursery mortality provides a valuable alternative for control of *H* parasuis, compared to traditional vaccination using commercial or autogenous products.

Keywords: swine, *Haemophilus parasuis*, control, vaccination, controlled exposure

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aemophilus parasuis is normally isolated from the nasal cavity, Ltonsils, and trachea of healthy pigs.^{1,2} This organism can potentially invade the host and cause severe systemic infection characterized by fibrinous polyserositis, arthritis, and meningitis. The factors involved in systemic invasion by *H parasuis* have not been clearly defined.³ The potential interaction between *H parasuis* and some viral agents, such as porcine reproductive and respiratory syndrome virus (PRRSV),4 pseudorabies,⁵ and porcine circovirus type 2 (PCV2),⁶ has been investigated. However, no direct association between any of these agents and *H parasuis* has been conclusively established. The relationships between Hparasuis virulence and serovar,⁷ genotype,⁸ and whole-cell protein profiles have also

been evaluated. Although virulent strains share similar genotype and protein profiles, no specific virulence factors have been described for *H parasuis*.

Antibiotic treatments, or vaccination with either commercial or autogenous products, may be used to control *H parasuis*. The effectiveness of autogenous vaccines against homologous challenge has been demonstrated. A lack of cross-protection between different *H parasuis* serovars and strains has also been reported. Commercial vaccines provide satisfactory homologous protection against *H parasuis* strains from the same serovar group. However, vaccination failures using commercially available products have also been demonstrated, especially when the *H parasuis* strains prevalent in the herd differ

from those included in the vaccine. In this case, autogenous vaccines may be used to reduce nursery mortality.¹⁷

Controlled exposure of 5-day-old pigs to a low dose of live, virulent *H parasuis* has been recently proposed as an alternative method to control nursery mortality. 18 This method is based on the hypothesis that early colonization of piglets with the prevalent strains of *H parasuis* in the presence of maternal immunity reduces the risk of systemic infection after weaning. This control measure has been reported to reduce morbidity¹⁹ due to Streptococcus suis and mortality 18 due to S suis and H parasuis systemic infection in affected nurseries. In the present study, control of H parasuis in a herd experiencing high nursery mortality was attempted using a commercial vaccine, an autogenous vaccine, or controlled exposure to live *H parasuis* in different cohorts.

SO, CP, RM: Department of Clinical and Population Sciences, College of Veterinary Medicine, University of Minnesota, St Paul, Minnesota.

Corresponding author: Dr Carlos Pijoan, University of Minnesota, College of Veterinary Medicine, Department of Clinical and Population Sciences, 1988 Fitch Ave #385, St Paul, MN 55108; Tel: 612-625-2245; Fax: 612-625-1210; E-mail: pijoa@@latc.umn.edu.

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Materials and methods

Multi-farm system

A multi-farm system (30,000 sows) experiencing high nursery mortality due to *H* parasuis infections 1 to 4 weeks after weaning was selected to evaluate vaccination and

controlled exposure as control measures. Tissue samples collected from nursery pigs in this system were positive by polymerase chain reaction (PCR) for PRRSV, PCV2, and swine influenza virus (SIV), and were negative by PCR for *Mycoplasma hyopneumoniae*.

Pigs were weaned at an average of 21 days of age and transferred to off-site nurseries. Each nursery contained eight rooms, which were filled with approximately 1000 pigs each. Pigs remained in the nursery for 7 weeks.

Experimental design

The mortality data (3.25%, SD 1.1%) obtained from the last nonvaccinated group of pigs housed in the nurseries used in this study were used to calculate the sample size needed to detect a 1% decrease in mortality, with $\alpha = .05$ and a power of 80%. Three experiments were performed. In Experiment 1, pigs were vaccinated with a commercial H parasuis vaccine. In Experiment 2, pigs were vaccinated with an autogenous vaccine containing two different strains. In Experiment 3, pigs were exposed to a low dose of live, virulent H parasuis (three different strains). Experiments were not concurrent, and were performed using different cohorts of pigs.

The experimental design used in each experiment is summarized in Table 1. In each experiment, a randomized complete block design was used, with barn as the blocking factor and room as the experimental unit. At the start of the trial, each barn was emptied, cleaned, and disinfected. One barn was filled each week for 5 consecutive weeks (Experiments 1 and 2) or 9 consecutive weeks (Experiment 3). Treatment was allocated to rooms within barns by randomly selecting the first room of the first barn to be used for a control or vaccinated group, and consecutive rooms were systematically allocated thereafter.

Mortality data, farm visits, and sample collecting and processing

Farm personnel were instructed to report mortality data at each nursery closeout. Mortality data used for comparison between controls and treated groups included pigs that died and pigs that were euthanized. Farm visits, for the purpose of determining the prevalent strains of *H parasuis* in the nursery, were completed before the beginning of Experiments 2 and 3, and during the

Table 1: Experimental design used to assess the effectiveness of vaccination and controlled exposure for control of nursery mortality due to *Haemophilus parasuis* infection

| Experiment | | Control measure | Timing | No. of rooms ¹ | | No. of replicates |
|------------|------------|------------------------------------|------------------------------------|---------------------------|---------|-------------------|
| # | Start date | | | Control | Treated | _ |
| 1 | Dec 2001 | Commercial vaccine ² | At weaning | 2 | 6 | 5 |
| 2 | Jul 2002 | Autogenous vaccine ³ | At processing and at weaning | 4 | 4 | 5 |
| 3 | Jan 2003 | Controlled exposure ⁴ | At processing | 2 | 2 | 9 |

- Each nursery barn contained eight rooms each with a capacity for 1000 pigs. One barn housing experimental groups was filled each week for 5 consecutive weeks (Experiments 1 and 2) or 9 consecutive weeks (Experiment 3).
- ² Ingelvac HP-1; Boehringer Ingelheim Vetmedica, Inc, St Joseph, Missouri. Pigs were vaccinated once intramuscularly at weaning (21 days of age).
- The autogenous vaccine (killed bacterin with the same adjuvant used in the commercial vaccine) included one representative *H parasuis* isolate from each group of prevalent strains (A and B) recovered from nursery pigs in Experiment 1. Pigs were vaccinated intramuscularly with a dose of 2 mL at processing (5 days of age) and at weaning (21 days of age).
- The inoculum included one representative *H parasuis* isolate from each group of prevalent strains (A, B, and C) recovered from nursery pigs in Experiment 2. Pigs were inoculated orally at processing (5 days of age) using a spray pump that delivered a 1-mL dose of bacterial suspension containing approximately 10⁵⁻⁶ colony forming units per mL.

fourth week after the first exposed pigs from Experiment 3 had entered the nursery (Table 1). Samples collected for detection of PRRSV, SIV, PCV2, and *M hyopneumoniae* were tested by PCR at the Veterinary Diagnostic Laboratory (University of Minnesota, St Paul, Minnesota). Samples collected for *H parasuis* isolation and detection by PCR were processed at the laboratory of Dr Pijoan (College of Veterinary Medicine, University of Minnesota, St Paul, Minnesota). *Haemophilus parasuis* fingerprinting and genetic analysis were also performed at Dr Pijoan's laboratory.

Vaccines

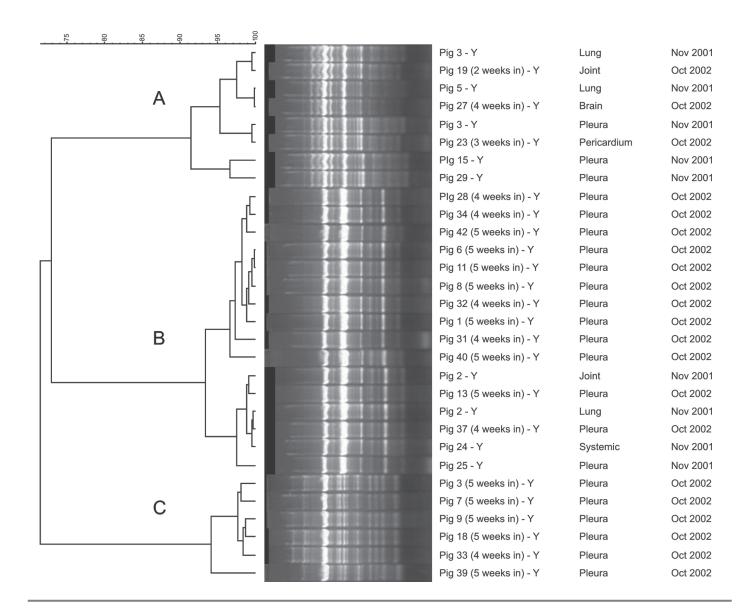
Pigs in Experiment 1 were vaccinated intramuscularly at weaning (21 days of age) using a single dose of a commercially available H parasuis bacterin (Ingelvac HP-1; Boehringer Ingelheim Vetmedica, Inc, St Joseph, Missouri). This vaccine is reported to protect against heterologous challenge. ²⁰

In order to select the *H parasuis* strains for inclusion in the autogenous vaccine (Experiment 2), 32 nursery pigs were necropsied, including 15 clinically affected pigs and 17 pigs found dead, and swabs for *H parasuis* isolation were collected from pleura, peri-

cardium, peritoneum, joints, and meninges, plus lung tissue. Swabs were cultured onto blood agar with a nurse Staphylococcus aureus streak, and plates were incubated at 37°C for 24 hours. Collected swabs were also tested by PCR21 to assess the prevalence of *H parasuis* systemic infection in the nursery. Haemophilus parasuis isolates were genotyped by enterobacterial repetitive intergenic consensus-based PCR (ERIC-PCR) as described previously. 8 Genomic fingerprints were analyzed using the BioNumerics software (Applied Maths, Kortrijk, Belgium), and one representative isolate from each prevalent group of H parasuis strains was included in the autogenous vaccine. Haemophilus parasuis isolates (one strain from genotype group A and one strain from genotype group B; Figure 1) were forwarded to Boehringer Ingelheim Vetmedica, Inc (St Joseph, Missouri), where the autogenous vaccine was produced with an adjuvant similar to that used in the commercial vaccine. Haemophilus parasuis was inactivated using 0.3% formalin. The vaccine was formulated to contain a minimum of 10⁸ of *H parasuis* organisms per mL. Pigs were vaccinated intramuscularly at

Figure 1: Cluster analysis by means of enterobacterial repetitive intergenic consensus-based PCR (ERIC-PCR) fingerprinting of 30 *Haemophilus parasuis* isolates recovered from affected nursery pigs in a production system experiencing high mortality due to *H parasuis* 1 to 4 weeks after weaning at 21 days of age. The scale indicates the percentage of genetic similarity. Columns (left to right) indicate isolate identification, site of isolation, and date of isolation. Three major clusters of prevalent strains are identified (A, B, and C).

Pearson correlation (Opt:1.00%) [0.0%-0.4%] [17.8%-79.0%] [99.8%-100.0%] ERIC-PCR ERIC-PCR



processing (5 days of age) and at weaning (21 days of age) using a 2-mL dose. Negative control pigs were not vaccinated.

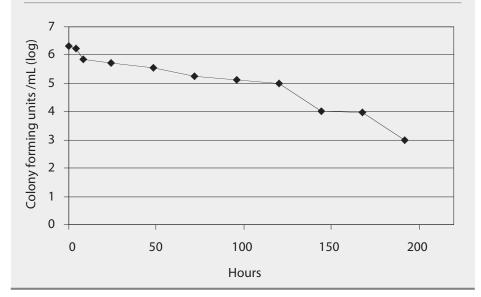
Inoculum preparation and administration

Forty-three animals were necropsied, including 41 clinically affected pigs and two pigs found dead. Pools of lung tissue obtained from these pigs were tested for PRRSV, PCV2, SIV, and *M hyopneumoniae* by PCR. Samples were collected and the

obtained *H parasuis* isolates were characterized as described. One representative *H parasuis* isolate from each group of prevalent strains (A, B, and C) was included in the inoculum used for controlled exposure (Figure 1). *Haemophilus parasuis* isolates were cultured onto chocolate agar and incubated in a candle jar at 37°C for 24 hours. A seed culture containing the prevalent *H parasuis* strains isolated from affected animals was prepared by harvesting the bacterial growth from chocolate agar plates

and suspending it in 10% sterile skim milk. The bacterial suspension was distributed in 1-mL aliquots and stored at -20°C until use. The final inoculum was prepared by diluting a 1-mL aliquot in 1 liter of sterile phosphate buffered saline (PBS).²² The inoculum was then transferred to plastic spray pumps and maintained at 4°C until use. A survivability test was performed in order to assess the viability of the inoculum after dilution in PBS. The diluted inoculum was maintained at 4°C and bacterial counts

Figure 2: Graph showing viable counts of the *Haemophilus parasuis* inoculum used in the controlled exposure of 5-day-old pigs. Counts were made at 0, 4, 8, 24, 48, 72, 96, 120, 144, 168, and 192 hours after dilution in phosphate buffered saline. The bacterial suspension was maintained at 4°C during and after dilution.



were performed at 0, 4, 8, 24, 48, 72, 96, 120, 144, 168, and 192 hours after dilution (Figure 2).

Pigs were inoculated with 10⁵⁻⁶ colonyforming units (CFU) per mL by the oral route. At processing, each piglet in the treatment group received an oral spray (1-mL dose) of the inoculum containing 10⁵⁻⁶ CFU per mL. Pigs in the control group were not exposed to *H parasuis*. The delivered amount of inoculum was validated by collecting the volume delivered by one spray in a microcentrifuge tube. All spray pumps were permanently regulated to deliver a 1-mL dose by using a pen marker. The same spray pumps were used throughout the whole experiment. Four weeks after the first group of inoculated pigs had been weaned, two or three pigs from each treated and control room were selected for euthanasia, on the basis of clinical signs characteristic of *H parasuis* systemic infection. Isolates from these pigs were cultured onto blood agar with a nurse streak of Saureus, and plates were incubated at 37°C for 24 hours. Swabs were tested by PCR for H parasuis. Haemophilus parasuis isolates were genotyped by ERIC-PCR and compared using the BioNumerics software.

Statistical analysis

The room (air space) was used as the experimental unit. A main effect ANOVA was performed in Statistica (StatSoft, Inc, Tulsa, Oklahoma), with percent mortality

at closeout as the dependent variable and room and treatment as the independent variables. The average mortality in control and treated rooms was compared.

Results

Isolation, PCR, and genotyping of *H parasuis*

Haemophilus parasuis isolation and PCR results are summarized in Table 2. Based on PCR results, prevalence of *H parasuis* systemic infection in the nursery was 50.0% at the first visit, 53.5% at the second visit,

and 35.3% at the third visit. Two prevalent genotypes (A and B) were identified among the isolates recovered from seven pigs in the first farm visit, and a third group of prevalent strains (genotype C) was identified among the isolates recovered from 20 pigs necropsied at the second farm visit. Isolates A and B were found to be still prevalent 1 year after the first isolation (Figure 1). At the third visit, when the first group of inoculated pigs had been in the nursery for 4 weeks, only strains A and B were recovered from clinically affected animals. Haemophilus parasuis was mainly isolated from nontreated control pigs (four of five isolates, compared to one isolate recovered from the joint of an inoculated pig).

In addition to *H parasuis*, *S suis* was isolated, during the first farm visit, from the lungs of six pigs found dead and from joints of two euthanized pigs. *Pasteurella multocida* (20 of 43 pigs) and *S suis* (six of 43 pigs) were also isolated from lung samples collected during the second farm visit. Pools of lung tissue collected during the second farm visit from pigs 2 weeks postweaning were PCR-negative for PRRSV, PCV2, SIV, and *M hyopneumoniae*. Pools from pigs 3 and 4 weeks postweaning were PCR-positive for PRRSV and PCV2. Pools from pigs 5 weeks postweaning were PCR-positive for PRRSV, PCV2, and SIV.

Experiment 1 (commercial vaccine)

Statistical analysis showed that percent mortality was normally distributed across all rooms, with an average of 5.1% (SD

Table 2: Haemophilus parasuis isolation and polymerase chain reaction (PCR) results for swabs collected from nursery pigs euthanized or found dead¹

| Date of farm | Number of pigs necropsied | | | Time in the nursery | No. of pigs positive (%) ² | | Prevalent genotypes |
|--------------------|------------------------------|------|-------|---------------------------|---------------------------------------|---------------|---------------------|
| visit | Euthanized | Died | Total | (weeks) | PCR | Isolation | |
| Nov 2001 | 15 | 17 | 32 | 2 – 6 | 16 (50.0) | 7 (21.88) | A, B |
| Oct 2002 | 41 | 2 | 43 | 2 – 5 | 23 (53.48) | 20 (46.51) | A, B, C |
| Mar 2003 | 17 | 0 | 17 | 3 – 4 | 6 (35.29) | 5 (29.41) | A, B |

- Each off-site nursery barn, containing eight rooms each with a capacity for 1000 pigs, was filled weekly with a cohort of pigs weaned at an average of 21 days of age. High nursery mortality due to *H parasuis* infections was occurring in this production system 1 to 4 weeks after weaning.
- Swabs collected from pleura, pericardium, peritoneum, joints, and meninges, plus lung tissue, were cultured for *H parasuis* isolation and tested by PCR. Pigs positive by isolation were also positive by PCR. Isolates were genotyped by enterobacterial repetitive intergenic consensus-based PCR.

1.74%). Average mortality was 4.8% (SD 1.17%) in nonvaccinated groups (n = 10) and 5.2% (SD 1.22%) in vaccinated groups (n = 30). Room was significantly associated with mortality (data not shown), while treatment was not.

Experiment 2 (autogenous vaccine)

As in Experiment 1, percent mortality appeared to be normally distributed across all rooms, with an average of 7.4% (SD 3.28%). Average mortality was 7.7 % (SD 2.02%) in nonvaccinated groups (n = 20) and 7.1% (SD 2.47%) in vaccinated groups (n = 20). Neither room nor treatment was significantly associated with mortality.

Experiment 3 (controlled exposure)

Across all rooms, percent mortality appeared to be normally distributed, with an average of 10.4% (SD 5.99%). Average mortality was 14.3% (SD 4.98%) in control groups (n = 18) and 6.4% (SD 1.79%) in treated groups (n = 18). Treatment was significantly associated with mortality (P < .001), while room was not.

Discussion

Results of this study showed that nursery mortality was not affected by vaccination using either commercial or autogenous vaccines, while mortality was lower in pigs exposed at 5 days of age to a low dose of live, virulent *H parasuis*. Although these three control measures were tested in the same herd, they were used in different populations of pigs at different time points. Considering these factors, care should be taken that each study be assessed individually.

The lack of cross-protection between the commercial vaccine strain and the prevalent *H parasuis* strains in the herd is a potential factor that might have influenced the lack of effectiveness of the selected vaccine. Other factors to be considered include incorrect timing of vaccination, interference by maternally-derived immunity, the dose used, and possibly a need for a booster vaccination, although the manufacturer's label directions were to use only one dose.

In the second trial, the failure of vaccination using the autogenous product was unexpected, since homologous protection has been reported to be effective. ^{10,17} Previous results obtained in field trials suggest that use of autogenous vaccines may be a viable alternative to control *H parasuis* when commercial products are not effective. ¹⁷ However, in the present study, it was demonstrated

that this may not be true for all swine herds. Again, several factors might have influenced the effectiveness of the autogenous vaccine, especially timing of vaccination and interference of maternal immunity. In the present study, pigs were vaccinated with the autogenous product at processing and at weaning due to the early onset of *H* parasuis systemic infection in the nursery (1 to 4 weeks after weaning). When pigs are initially vaccinated at such an early age, there is always a concern regarding interference of maternal immunity in the development of the active immune response induced by the vaccine. However, some studies have demonstrated that maternal immunity may not always interfere with the immune response. Solano-Aguilar et al¹⁵ tested a commercial vaccine containing H parasuis serovars 4 and 5, using two doses, and showed that vaccinated pigs born to vaccinated gilts were protected against challenge with a virulent *H parasuis* strain, whereas some vaccinated pigs born to nonvaccinated gilts developed central nervous system signs and lameness. Baumann and Bilkei¹⁶ also demonstrated that vaccination of sows and their piglets resulted in protection against homologous challenge. Further studies are necessary to better characterize the potential interference of maternal antibodies in pig vaccination. Another factor that might have influenced the outcome observed in Experiment 2 was the nonrandomized distribution of experimental groups in rooms within barns. This difference may have impaired the observation of a greater effectiveness of the autogenous vaccine in the treatment group.

Another potential factor to be considered is that the autogenous vaccine was missing one of the prevalent genotypes later identified in the nursery (type C). Genotyping results showed that strains A and B, which were isolated in the previous year, were still prevalent in the herd. A third prevalent strain (C) was identified after the second sampling. This strain was not included in the autogenous vaccine. Strain C was later included in the inoculum used for controlled exposure. A general increase in mortality was observed in the second experiment compared with the first experiment. This increase in mortality coincided with a PRRS outbreak in the system. Mortality was even higher in the control group after the third experiment compared with controls in the first and second experiments.

This time, SIV was identified as the main agent co-infecting nursery pigs. However, the association between PRRS or SIV and *H parasuis* co-infection in the nursery has not been established.

Regarding controlled exposure, Pijoan et al²³ hypothesized that early mucosal colonization of piglets with the herd's prevalent strains of *H parasuis* while they are still protected by maternal immunity might reduce the risk of systemic infection after weaning. Following this hypothesis, it would be desirable that a large number of pigs be exposed to these potentially virulent strains before they become susceptible (ie, with waning of maternal antibody levels). Some practices currently used in modern swine production, such as early weaning and three-site production systems, may reduce exposure to these virulent strains, or spread of infection among pigs. Kirkwood et al²⁴ demonstrated that levels of colonization by *H parasuis* appear to be influenced by weaning age. Their study showed that levels of colonization were lower in pigs weaned at 14 days than in pigs weaned at 28 days.

The mechanisms involved in development of protective immunity after controlled exposure to *H parasuis* have not been clearly defined. Nielsen²⁵ demonstrated that exposure of specific-pathogen-free pigs to an aerosol containing live, apathogenic *H parasuis* strains resulted in development of circulating antibodies, and protected pigs against heterologous challenge with a virulent strain. However, further studies are necessary to better characterize the mechanisms involved in development of protective immunity by pigs exposed to live, virulent *H parasuis*.

In the present study, nursery mortality was significantly lower in treated groups after controlled exposure than in untreated control groups. We have previously reported that after colonization of piglets with a low dose of live, virulent H parasuis, nursery mortality was 36.6% lower in the exposed group than in the control group. 18 Four factors may have improved the results of the colonization trial in the present study compared with our previous report. The sample size and number of replicates (8000 pigs per group, five to nine replicates) used in the present study were considerably larger than those in the previous report (50 pigs per group, two replicates). The herd

used in the present study had a greater prevalence of systemic infection and mortality due to *H parasuis*. The dose of inoculum used in 5-day old pigs was higher in the present study (1 × 10^{5-6} CFU per mL) than in the previous study (7 × 10^3 CFU per mL). The allocation of treated and control groups in the nursery, as well as the experimental unit used for statistical analysis, differed between studies. In the present study, treated and control groups were allocated to different rooms in a single barn, and room, or air space, was considered the experimental unit. In our previous study, all groups were allocated to different pens in the same nursery barn (ie, the same air space), and the pig was considered the experimental unit.

Results obtained in the present study demonstrated that nursery mortality may be significantly reduced after early exposure of pigs to live, virulent *H parasuis*, compared with the results of using autogenous and commercial vaccines. Controlled exposure has several advantages compared with traditional vaccination, including lower cost and reduction of workload. Furthermore, timing does not seem to be an issue with controlled exposure, whereas maternal immunity may interfere with pig vaccination. There are some concerns regarding the safety of this method. The interaction between PRRS virus and H parasuis has not been scientifically demonstrated. However, field experiences suggest that these organisms may co-infect nursery pigs, resulting in increased mortality compared with either pathogen alone. It would appear to be counterindicated to inoculate pigs with live, virulent H parasuis if PRRS virus infection is active in the sow farm. The vaccination results obtained in this study suggest that the use of commercial and autogenous vaccines to control H parasuis infections must be critically evaluated.

Implications

- Controlled exposure may be an alternative to other methods for control of *H parasuis* systemic infection in the nursery.
- Under the conditions of this study, controlled exposure was more effective than use of autogenous or commercial vaccines in reducing nursery mortality due to *H parasuis* systemic infection.
- Although the interaction of PRRSV and *H parasuis* has not been scientifically proven, as a safety

measure, nursery pigs should not be exposed to live, virulent *H parasuis* when there is active PRRSV infection in the sow herd.

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